

Sulfonomethyl analogues of aldoses-2-ulosonic acids. Synthesis of a new sialyl Lewis X analogue

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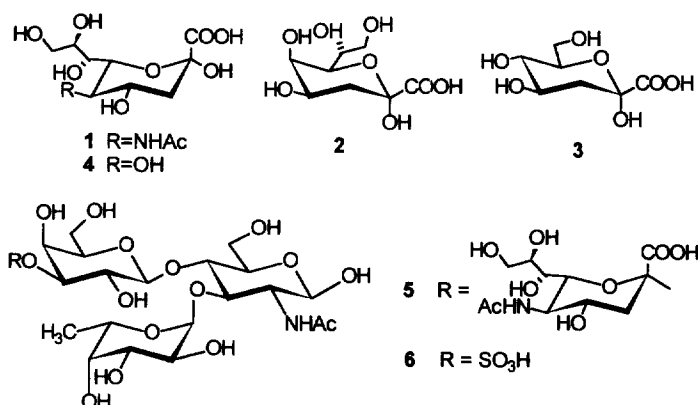
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Abstract: Sulfonomethyl derivatives of aldoses-2-ulosonic acids have been synthesized by addition of the ethyl methanesulfonate carbanion to aldolactones. A sulfonfylated mimic molecule of the sialyl Lewis X tetrasaccharide has been prepared by using a new sulfonomethyl ulosonic acid analogue.

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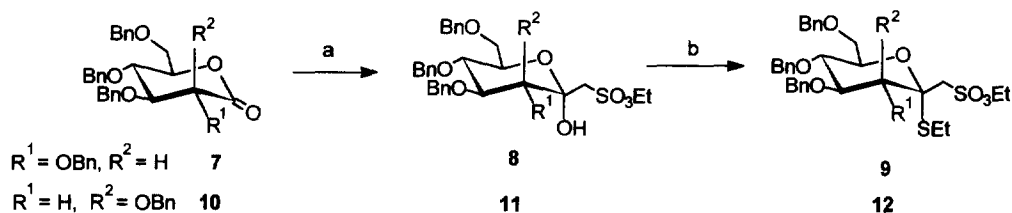
Aldoses-2-ulosonic acids such as *N*-acetylneuraminic acid [1] (**1**), 3-deoxy-*D*-manno-2-octulosonic acid [2] (KDO, **2**), 3-deoxy-*D*-arabino-2-heptulosonic acid [3] (**3**) and 3-deoxy-*D*-glycero-*D*-galacto-nonulosonic acid [4] (KDN, **4**) play important roles in various biological processes as mediators of cell-viruses or cell-cell recognition [5-7], intermediates of the shikimic acid biosynthesis [3], and constituents of bacterial cell walls [2], etc. The sialyl Lewis X tetrasaccharide (sLe^x) [8] (**5**) - containing *N*-acetylneuraminic acid - is a ligand for E- and P-selectin, therefore is a key interactive molecule of inflammatory reactions and metastasis.



Scheme 1

Mimics of 1-5 can be used as inhibitors of enzymes and recognition phenomena [8-10]. The charge of the carboxylic group in 2-ulosonic acids is presumably essential for binding to biomolecules, therefore substitution of carboxyls for another charged moieties could produce bioactive surrogates of 1-5. While phosphonic acid analogues of 1 are known [11,12], sulfonic acid variants have not been synthesized to date, although several sulfo sugars, having mainly a 6-sulfonic acid moiety, are known [13-20]. Taking into consideration that sulfonic acids are stronger acids than their carboxylic counterparts it can be assumed that sulfonate analogues of 2-ulosonates could bind effectively to the bioreceptors. This idea seems to be confirmed by the fact that the sulfated Lewis X trisaccharide 6 does exist, as a natural analogue of sLe^x, and shows superior binding to E-selectin [21]. This observation has stimulated significant interest in the synthesis of sulfated Lewis^x derivatives [21-25]. In this paper we wish to present preliminary results on the preparation of sulfonomethyl analogues of aldoses-2-ulosonic acids and of a sLe^x tetrasaccharide.

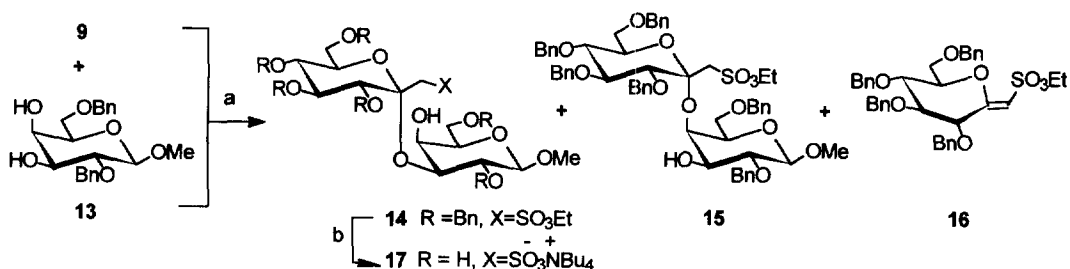
For the synthesis of the model compound 8, the D-gluconolactone derivative 7 [26] was reacted with the ethyl methanesulfonate anion generated with *n*-butyllithium. Upon nucleophilic addition of the sulfonate ester carbanion to the lactone carbonyl the 1-ethylsulfonyl-D-hept-2-ulose 8 was obtained in α -anomeric form. The anomeric configuration was determined on the basis of the NMR C1-H3 three-bond coupling constant [27], that depends on the dihedral angle in a manner similar to ³J_{H,H}.



Scheme 2: a) CH₃SO₃Et, *n*-BuLi, THF, -70 °C, ~90%; b) EtSH, BF₃Et₂O, CH₂Cl₂, ~95%

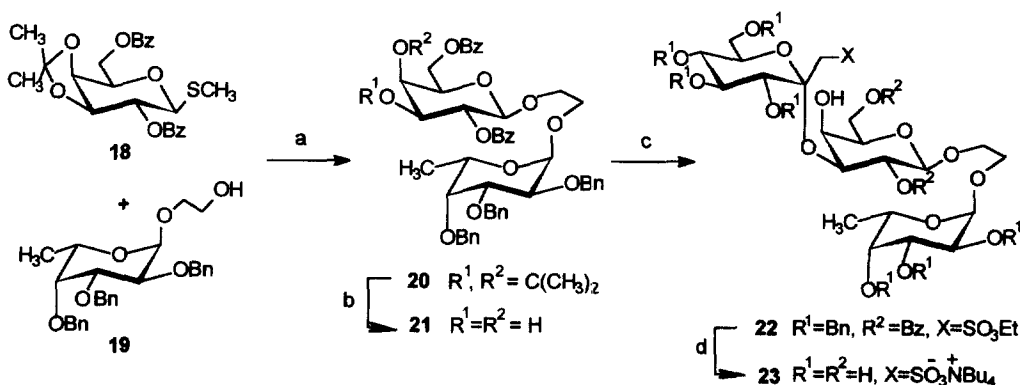
Reaction of 8 with ethanethiol in the presence of Lewis acid resulted in the formation of the α -thioglycoside 9. The same reaction sequence starting from the D-mannonolactone 10 [28] furnished the thioglycoside 12.

Since our main goal was the substitution of *N*-acetylneuraminic acid with sulfonomethyl derivatives in sialyl Lewis^x tetrasaccharide analogues we first investigated the regioselective glycosylation of the diol 13 [29] at position 3. Glycosylation of 13 with 9 using *N*-iodosuccinimide – trifluoromethanesulfonic acid (NIS-TfOH) activation afforded a separable 3:1 mixture of the regioisomeric disaccharides 14 and 15. Formation of the elimination product 16 was also observed with an isolated yield of 13%. The protecting groups were removed from 14 *via* nucleophilic attack by bromide and subsequent catalytic hydrogenation resulted in the tetrabutylammonium salt 17.



Scheme 3: a) NIS, TfOH, CH₂Cl₂, -50 °C, 55% of 14, 17% of 15, 13% of 16; b) Bu₄NBr, CH₃CN, reflux; EtOH, Pd(C)/H₂, 87% (2 steps)

For the synthesis of the Le^x trisaccharide mimic **19** was glycosylated with the donor **18** [30] using methyl triflate activation to give **20** which, following deisopropylideneation, may serve as the aglycon for further glycosylation. The thioglycoside donor **9** was coupled with the diol **21** using NIS-TfOH activation, and regioselective formation of **22** was observed due to the reduced reactivity of **21**, but the yield was rather low (~35%). The reason for this is the reduced reactivity of the aglycon, and so that an up to 50% increase of the elimination product **16** from the donor **9**. By means of a three-step deprotection procedure **22** was converted into the sulfonic-acid type mimic **23** of the sLe^x tetrasaccharide.



Scheme 4: a) MeOTf, CH₂Cl₂ 86%; b) HCl, MeOH, 40 °C, 97%; c) **21**+**9**, NIS, TfOH, CH₂Cl₂ -50 °C, 35% of **22**, 50% of **16**; d) NaOMe, MeOH; Bu₄NBr, CH₃CN, reflux; EtOH, Pd(C)/H₂, 84% (3 steps)

The synthesis of further sulfonomethyl analogues of aldose-2-ulosonic acids and their introduction into oligosaccharides are under way in our laboratory.

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All compounds gave satisfactory spectroscopic data. Selected spectroscopic and physical data are the following: compound **9**: [α]_D +59.94 (c 0.61, CHCl₃), ¹³C NMR (125 MHz, benzene): δ 89.7 (C-2), 84.7 (C-4), 80.7 (C-3), 79.8 (C-5), 75.0 (C-6), 69.3 (C-7), 66.8 (SO₃CH₂CH₃), 56.7 (C-1, J_{H3,C1} 2.7 Hz), 20.1 (SCH₂CH₃), 15.1 (SO₃CH₂CH₃), 13.9 (SCH₂CH₃). Compound **12**: [α]_D +11.15 (c 0.26, CHCl₃), ¹³C NMR (90 MHz, CDCl₃): δ 86.6 (C-2), 81.8, 76.8, 74.5, 73.4 (C-3, -4, -5, -6), 69.1 (C-7), 66.3 (SO₃CH₂CH₃), 54.0 (C-1), 20.6 (SCH₂CH₃), 14.9 (SO₃CH₂CH₃), 13.5 (SCH₂CH₃). Compound **15**: [α]_D +34.86 (c 0.72, CHCl₃), ¹³C NMR (90 MHz, CDCl₃): δ 105.2 (C-1), 99.2 (C-2'), 82.6, 80.5, 80.1, 78.4 (C-3', C-5', C-2, C-4), 75.4, 75.3, 75.2, 74.6, 73.4, 72.6 (6 OCH₂Ph), 73.7, 73.4, 72.8, 72.6 (C-4', C-6', C-3, C-5), 68.8, 68.6, 67.8 (C-6, C-7', SO₃CH₂CH₃), 57.2 (OCH₃), 51.8 (C-1, J_{H3,C1} <1 Hz), 15.0 (SO₃CH₂CH₃). Compound **16**: ¹³C NMR (90 MHz, CDCl₃) 165.4 (C-2), 107.6 (C-1), 85.7, 81.9, 80.3, 80.1 (C-3, -4, -5, -6), 71.3 (C-7), 70.2 (SO₃CH₂CH₃), 18.3 (SO₃CH₂CH₃). Compound **17**: [α]_D +44.27 (c 1.10, H₂O), ¹³C NMR (90 MHz, D₂O): δ 106.5 (C-1), 102.7 (C-2'), 77.2 (C-3), 75.8, 75.6, 75.3 (C-3', C-5', C-2, C-4), 72.3, 71.9, 71.5 (C-4', C-6', C-5), 63.7, 63.4 (C-6, C-7'), 60.8 (N⁻-CH₂-), 59.8 (OCH₃), 56.9 (C-1', J_{H3',C1'} 2.0 Hz), 25.8, 21.8 (N⁻-CH₂-CH₂-CH₂-CH₃), 15.4 (N⁻-CH₂-CH₂-CH₂-CH₃). Compound **21**: [α]_D -32.94 (0.36, CHCl₃), ¹³C NMR (90 MHz, CDCl₃) δ 167.1, 166.4 (2 CO), 100.4 (C-1-Fucp), 97.9 (C-1-Galp), 62.9 (C-6-Galp), 67.5, 73.0 (CH₂-CH₂). Compound **22**: [α]_D +22.58 (c 0.17, CHCl₃), ¹³C NMR (125 MHz, benzene): δ 166.2, 165.3 (2 CO), 100.6 (C-1), 100.3 (C-2''), 98.1 (C-1'), 64.0 (C-6'), 67.5, 67.7, 68.6 (CH₂-CH₂, C-7'', SO₃CH₂CH₃), 53.9 (C-1'', J_{H3'',C1''} <1 Hz), 16.9 (CH₃-Fucp), 15.1 (SO₃CH₂CH₃). ESI +Q1MS: M⁺Na⁺ 1515.7. Compound **23**: ¹³C NMR (125 MHz, D₂O): δ 105.7 (C-1'), 101.7 (C-1), 75.8 (C-6''), 75.3 (C-4''), 74.9 (C-3''), 72.0 (C-5''), 71.7, 69.8 (-CH₂-CH₂-), 63.7 (C-7''), 63.5 (C-6''), 59.1 (C-1''), 18.0 (C-6); ¹H NMR (500 MHz, D₂O): δ 4.9 (1 H, d, H-1, J_{1,2} 4.5 Hz), 4.5 (1 H, d, H-1', J_{1',2'} 8.8 Hz), 3.4, 3.5 (2 H, 2 d, H-1_a'', H-1_b'', J_{gem} 14 Hz).

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